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Synthesis and Electrochemical Characterization of TiO₂/g-C₃N₅ Coated 316L Stainless Steel for Orthopedic Applications

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Abstract: This study investigates the effect of varying amounts of nitrogen-rich carbon nitride (g-C₃N₅) incorporated into titanium dioxide (TiO₂) coatings on 316L stainless steel (316LSS). The TiO₂/g-C₃N₅ coatings were tested in simulated body fluid (SBF) to assess their performance for orthopedic applications. TiO₂ was prepared using the sol-gel method, while g-C₃N₅ was synthesized through thermal polymerisation. The crystal structure, purity, and chemical composition of the TiO₂/g-C₃N₅ (TiCN) composites were confirmed using X-ray diffraction (XRD), Fourier transform infrared (FTIR) spectroscopy, and Raman analysis. The surface morphology of the coated samples was characterised using Scanning Electron Microscopy (SEM). In contrast, surface roughness was measured with Atomic Force Microscopy (AFM), revealing a porous film with an average particle size of 25 to 100 nm was coated over 316LSS. A fourfold increase in corrosion resistance was evaluated through Open circuit potential (OCP), Potentiodynamic polarisation, and Electrochemical impedance spectroscopy (EIS). The in vitro test revealed the enhanced growth of a hydroxyapatite layer on the coated TiCN. The elemental composition of calcium and phosphate ions present in the hydroxyapatite (HAP) deposition was confirmed using Raman spectroscopy. The results suggest that the TiCN coated 316LSS was a promising material for biomedical applications.

Keywords: Biomaterials, Corrosion, Graphitic carbon nitrite (g-C₃N₅), Titanium dioxide, Stimulated body fluid

1. Introduction

Metal implants are significant in biomedical applications due to their superior corrosion resistance and mechanical properties under physiological conditions [1, 2]. Surgical grade 316L stainless steel receives remarkable attention in the field of biomedical implants due to its corrosion resistance, affordability, manufacturability, longevity, and widespread availability [3]. The metallic ions like Ni²⁺, Cr³⁺, and Cr⁶⁺ are released during the oxidation and contribute to the loosening of prosthetic devices by having localised adverse effects on the implanted area [4, 5]. Corrosion prevention of 316LSS implants is important because it can adversely affect both the biocompatibility and mechanical strength of the implants. As a result, optimising the corrosion resistance and minimising ion release of 316LSS implants can be effectively achieved by surface modification with bioactive coatings, which promote cellular adhesion and bone integration [6]. To permanently protect its surfaces, an ideal biomaterial needs to have a robust framework with features like biocompatibility and improved corrosion resistance properties [7]. The use of various compounds enhances

the biocompatibility and corrosion resistance. There have already been numerous attempts in this direction. Bioimplant materials have been subjected to coatings with a range of metal oxides, including TiO₂ [8, 9] ZrO₂ [10], SiO₂ [11], and Al₂O₃ [12]. Among these, titania (TiO₂) is used in the biomedical industry due to its high fracture resistance, durability, antibacterial activity and exceptional biocompatibility [13]. However, titanium dioxide (TiO₂) possesses certain inherent limitations, including poor mechanical properties of individual ceramic biomaterials that limit their performance in high load-bearing capacity [14]. Madan Kumar and Rajendran investigated the electrochemical and biological behavior of polypyrrole/titanium dioxide ceramic nanocomposite coatings on 316LSS and demonstrated that it is a promising material for orthopedic implants. These coatings reduce the corrosion rate and facilitate enhanced cell adhesion and proliferation [15]. K.T. Oh and Y. S. Park demonstrated that super austenitic stainless steel (SASS) coated with hydroxyapatite (HA) by plasma spraying provides excellent corrosion resistance and mechanical properties [16].

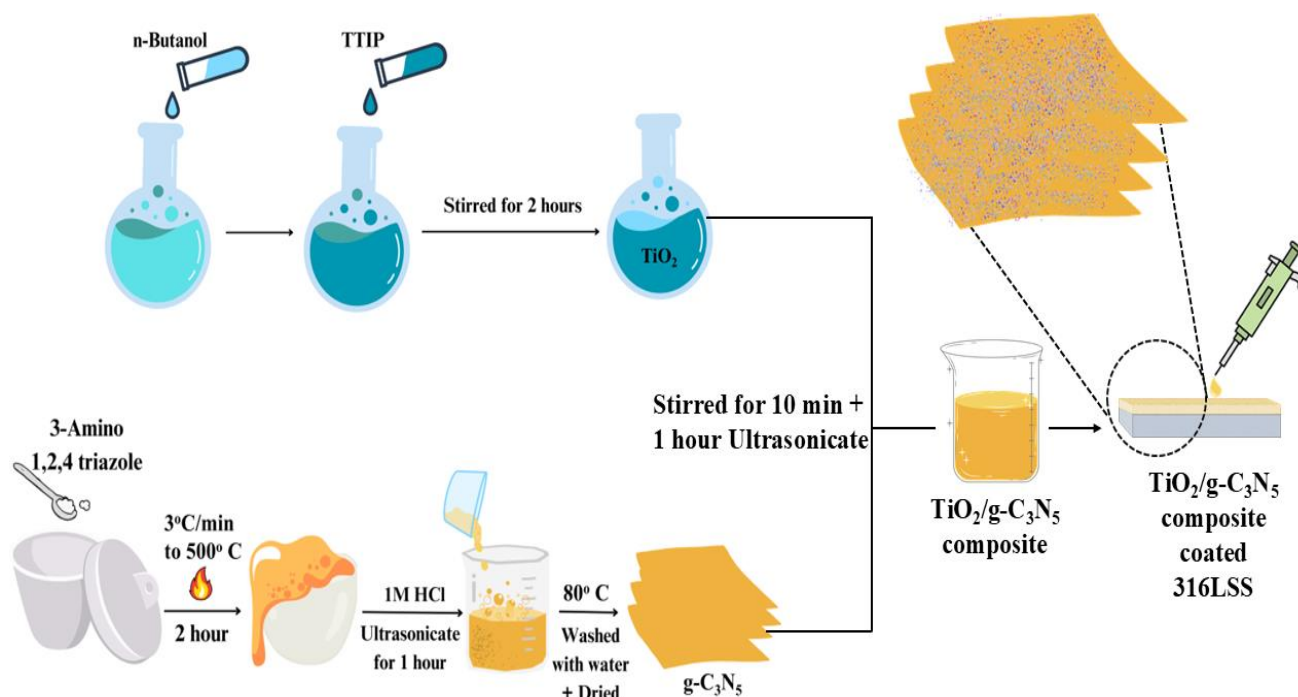


Figure 1. Schematic illustration for the formation of $\text{TiO}_2/\text{g-C}_3\text{N}_5$ Composites coating

Dai *et al.* reported that $\text{Al}_2\text{O}_3\text{-TiO}_2$ coatings with 15.56% titanium content visibly improved the corrosion resistance of 316LSS in chloride environments [17]. Garrido *et al.* showed that incorporating bioactive glass 45S5 with PEEK through cold gas spray improved the wear resistance and the integration between bone tissue and implant material [18]. TiO_2 -based coatings for biomedical implants emphasize their corrosion resistance, biocompatibility, and potential for surface modification to enhance performance. However, the limitations of pure TiO_2 , such as suboptimal mechanical stability and susceptibility to long-term degradation, highlight the need for composite materials. Therefore, to overcome these challenges, a well-established approach involves the preparation of nanocomposite coatings, which offer superior corrosion resistance compared to single-layer coatings. The success of an implant depends on the stability of its coating, which provides good biocompatibility. Using nanocomposites for surface modification is an effective strategy to improve implant performance [19-30].

$\text{g-C}_3\text{N}_5$, a graphitic carbon nitride material, has garnered attention in various fields, including photocatalysis, energy storage, and environmental remediation, due to its exceptional stability, chemical inertness, and tunable electronic properties. Its unique structure and synergistic potential when combined with other materials, such as TiO_2 , have been shown to enhance photocatalytic activity and mechanical properties. However, its application in biomedical coatings, particularly for improving corrosion resistance and biocompatibility in implantable devices, remains unexplored. Adding polymeric mesoporous graphitic carbon nitride ($\text{g-C}_3\text{N}_5$) has garnered significant attention

among the various methods for improving titania. $\text{g-C}_3\text{N}_5$ was chosen for its exceptional biocompatibility, high surface area, ordered porosity, stable chemical characteristics, biological activity and corrosion resistance. $\text{g-C}_3\text{N}_5$ offers a high nitrogen content and demonstrates remarkable properties across a wide range of applications 31-36. $\text{g-C}_3\text{N}_5$ is a new material, and there is limited review papers focus on understanding the photocatalytic applications of $\text{g-C}_3\text{N}_5$ compared to the $\text{g-C}_3\text{N}_4$. To date, no research has been done on the use of $\text{g-C}_3\text{N}_5$ in orthopedic applications. This highlights the need to explore its potential in this area, where its properties could offer new possibilities [37, 38]. As a result, $\text{g-C}_3\text{N}_5$ was successfully incorporated into TiO_2 for further investigation. There is no previous report of utilising 316LSS alloy coated with titanium dioxide and nitrogen-rich graphitic carbon nitride for biomedical applications. This study provides precise insights into the electrochemical behavior of materials and valuable information for developing effective corrosion-resistant coatings. Notably, applying substantial $\text{TiO}_2/\text{g-C}_3\text{N}_5$ coatings leads to marked improvements, enhancing biocompatibility and corrosion resistance in Simulated Body Fluid (SBF) solution. To date, no study has explored the use of $\text{g-C}_3\text{N}_5$ in combination with TiO_2 for developing corrosion-resistant coatings on 316LSS implants. Furthermore, there are no prior reports on utilizing this nanocomposite for orthopedic applications, despite the potential advantages of incorporating $\text{g-C}_3\text{N}_5$ into TiO_2 to enhance its properties. This study aims to develop and characterize $\text{TiO}_2/\text{g-C}_3\text{N}_5$ nanocomposite coatings for 316LSS implants, evaluate their electrochemical behavior, and assess their corrosion resistance in Simulated Body Fluid (SBF). Additionally, the

biocompatibility of the coatings will be assessed to determine their suitability for orthopedic applications. By addressing these objectives, this research seeks to advance the development of innovative corrosion-resistant coatings and establish g-C₃N₅ as a valuable material for biomedical applications.

2. Experimental

2.1 Materials

N-butanol (C₄H₁₀O) from RANKEM 99.0%, Titanium tetra isopropoxide (Ti (OCH (CH₃)₂)₄) from Sigma Aldrich (99.99 % purity), 3-Amino 1, 2, 4 triazole (C₂H₄N₄)-TCl, Hydrochloric acid solution (HCl)-Qualigens, Acetone (CH₃)₂CO, Ethanol (C₂H₅OH)-analytical grade reagents. Double distilled water was used throughout the experiment.

2.2 Preparation of 316LSS alloy sample

The chemical composition of 316LSS is shown in Table 1 [39]. The samples were fabricated from 316LSS and cut into 10 mm × 20 mm × 2 mm pieces. 80 to 2000 SiC paper was used to polish the metal at speeds between 200 and 900 rpm. After that, diamond paste was applied to provide a uniform, smooth surface texture. The sample was then cleaned in an ultrasonic bath using acetone, ethanol and distilled water. Finally, the polished 316LSS samples were used for further studies.

Table 1. Composition of the Simulated Body Fluid (SBF) solution used in the study, including concentrations of individual components and their respective molarities.

Order	Reagent	Amount (g)
1.	NaCl	12.052
2.	NaHCO ₃	0.535
3.	KCl	0.337
4.	K ₂ HPO ₄ .3H ₂ O	0.346
5.	MgCl ₂ .6H ₂ O	0.477
6.	1.0 M – HCl	58.5ml
7.	CaCl ₂	0.438
8.	Na ₂ SO ₄	0.108
9.	Tris (hydroxymethyl) aminomethane	9.177

2.3 Coating preparation

A mixture of titanium tetra isopropoxide and n-butanol was stirred under constant magnetic stirring for two hours to obtain a TiO₂ solution. To prepare g-C₃N₅ powder, 3-Amino 1, 2, 4 triazole was sintered at 500°C

for two hours. A foam-like powder was obtained and ground with a mortar. The powder was mixed with a 1M hydrochloric acid solution and placed in an ultrasonic cleaner. It was then sonicated for an hour. The powder was then repeatedly washed with deionised water. It was kept overnight in a hot air oven at 80°C. Different weights (0.01g, 0.03g, 0.05, 0.07g and 0.09g) of g-C₃N₅ were added to the prepared TiO₂ sol and stirred for 10 min. The suspension was subjected to ultrasonication for one hour. TiO₂/g-C₃N₅ (TiCN) sol was coated onto 316LSS at 4000 rpm for 20 seconds using a SPIN NXG-P1AC spin coater. The samples were sintered in a muffle furnace at 400°C for one hour after the coating process. The sintered samples were then collected and used for further investigation. The sintered samples were named as TiCN (0.01), TiCN (0.03), TiCN (0.05), TiCN (0.07) and TiCN (0.09) for further analysis.

2.4 Characterisation of sample

X-ray powder diffraction (XRD) spectra were collected utilising a Bruker D8 (Bruker AXS) diffractometer outfitted with a placement-sensitive detector. A CuKα lamp served as the irradiation source, emitting radiation with a wavelength of λ = 1.54Å. The crystal structure of the pure TiO₂, g-C₃N₅ and TiCN composite samples were analyzed. Samples were measured in the presence of infrared radiation, and chemical properties were observed using FTIR spectra in the 400–4000 cm⁻¹ range. WITec ALPHA300 Raman spectroscopy was used to obtain the Raman spectra. An air-cooled laser was the exciting source, emitting radiation at 532 nm. Atomic force microscopy (AFM) (MFP-3D Origin Asylum Research AFM) Oxford instrument was used to measure the surface topography of the TiCN Coated 316LSS, and the surface morphology of the film was observed by scanning electron microscopy (SEM) using CARL ZEISS instrument at an acceleration voltage of 10kV.

2.5 Electrochemical Evaluation

Electrochemical measurements were carried out using a standard three-electrode cell. In this configuration, a saturated calomel electrode (SCE) served as the reference electrode, a platinum electrode as the counter electrode, and the test material as the working electrode. Simulated body fluid (SBF) was used as the electrolyte. Polarization measurements were conducted from cathodic to anodic directions at a scan rate of 1 mV/s. Corrosion rate was calculated using the tafel extrapolation method. The impedance measurements were taken in a frequency range of 100 kHz to 0.1 Hz, with an amplitude of 10 mV. Electrochemical behavior and corrosion resistance tests were conducted with seven 316LSS samples per condition, each measured in triplicate for reliability and statistical significance. These experiments were performed using a (BIOLOGIC VSP 300) potentiostat

operated by a personal computer with (EC Lab version 8.5) software.

2.6 In vitro Studies

In vitro studies using SBF are widely conducted to evaluate the biocompatibility of materials for biomedical applications [40-42]. Coated samples were soaked in an SBF solution for 14 days to observe hydroxyapatite growth. SBF was prepared following the procedure described in the referenced article [43].

3. Result and Discussion

3.1 X-Ray Diffraction Analysis

X-ray diffraction (XRD) analysis was performed to characterise the crystalline phases of the synthesized sample. As shown in Figure. 2, both TiO_2 and $\text{g-C}_3\text{N}_5$ exhibited well-defined single-phase crystal structures. In the case of TiO_2 , the XRD pattern showed characteristic peaks at 25.2° , 37.5° , 47.9° , 53.7° , 54.8° , and 62.5° , corresponding to the (101), (004), (200), (105), (211), and (204) planes, which are indicative of the anatase phase [44]. For as-synthesized $\text{g-C}_3\text{N}_5$, the XRD pattern displayed a prominent peak at 27.6° , corresponding to the (002) plane, associated with the tighter packing of carbon nitride layers, consistent with JCPDS card no. 87-1526 [45,46]. Additionally, a broadened (002) peak with decreased intensity was observed for acid-treated $\text{g-C}_3\text{N}_5$ nanosheets, indicating exfoliation of the bulk material into fewer-layer nanosheets. This suggests that the bulk $\text{g-C}_3\text{N}_5$ was successfully exfoliated into smaller sheets. The TiCN composite displayed similar diffraction peaks as pure TiO_2 , suggesting that the crystal phase of TiO_2 remained unchanged in the presence of $\text{g-C}_3\text{N}_5$. However, slight variations in peak positions were observed in the composite, and some peaks showed

increased intensity, indicating intricate packing interactions between $\text{g-C}_3\text{N}_5$ and TiO_2 . Due to the relatively low amount of $\text{g-C}_3\text{N}_5$ in the TiCN composite, the prominent diffraction peaks for $\text{g-C}_3\text{N}_5$ were not observed [47]. No additional phases were detected beyond TiO_2 and $\text{g-C}_3\text{N}_5$, confirming the purity of the TiCN compositions.

3.2 FT-IR Analysis

FTIR analysis was used to investigate the elements in TiO_2 , $\text{g-C}_3\text{N}_5$, and TiCN samples are shown in Figure 3. The TiO_2 sample shows a broad band between 500 and 900 cm^{-1} , which may result from the formation of Ti-O and Ti-O-Ti bonds as the titanium dioxide. This broadening might also be due to the titanium dioxide film caused by carbon or hydroxyl groups in the Ti-O system [48, 49]. The $\text{g-C}_3\text{N}_5$ sample results in a peak at around 3100 cm^{-1} originates from the stretching vibration of $-\text{NH}_2$ and $-\text{OH}$ groups from water molecules adsorbed on the surface. The band between 1650 and 1250 cm^{-1} is due to the typical stretching of heterocyclic C=N units, while the peak at about 802 cm^{-1} represents the breathing vibration of triazine units. The peak near 1500 cm^{-1} , slightly shifted to a lower wavenumber and broadened compared to $\text{g-C}_3\text{N}_5$, suggests the formation of protonated $\text{g-C}_3\text{N}_5$, likely due to hydrolysis of C_3N_5 after acidification. This change also leads to the appearance of a C-O stretching peak at 1080 cm^{-1} . The broadened absorption peaks between 3500 and 3000 cm^{-1} for $-\text{NH}-$ or $-\text{NH}_2$ groups indicate that hydrogen bonds involving $-\text{NH}-/-\text{NH}_2$ were distorted [50, 51]. In the TiCN composites, the bands between 500 and 900 cm^{-1} are characteristic of TiO_2 , and the peaks between 3600 and 3800 cm^{-1} correspond to $\text{g-C}_3\text{N}_5$, confirming the interaction between TiO_2 and $\text{g-C}_3\text{N}_5$ in the TiCN composite.

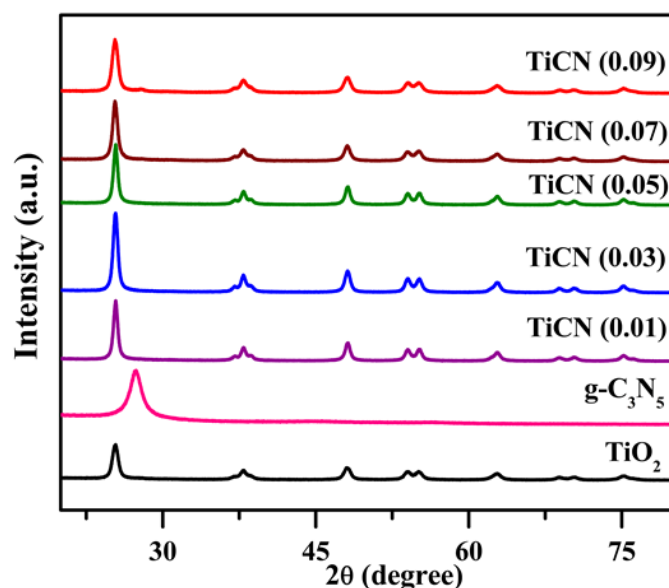


Figure 2. XRD patterns showing the structural and crystalline characteristics of the sample and their composites

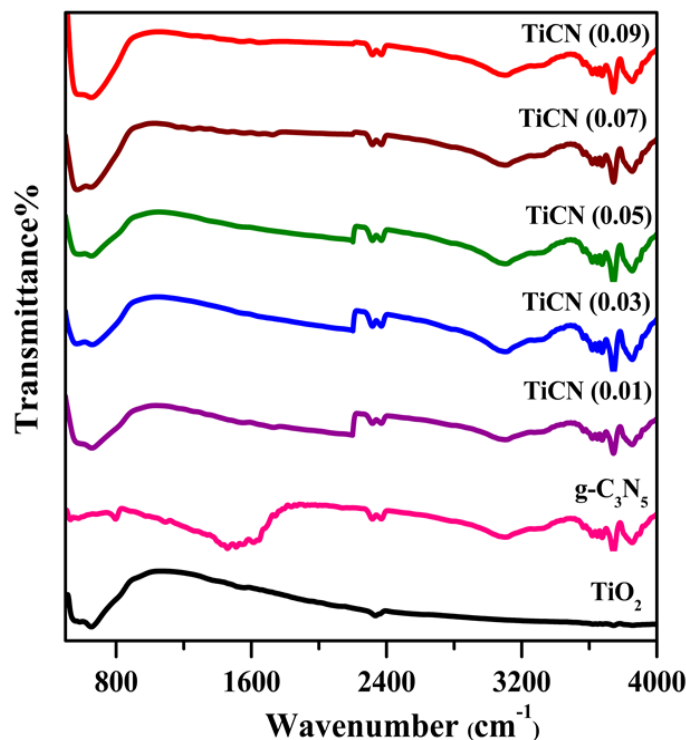


Figure 3. FTIR spectra showing characteristic absorption peaks of TiO_2 , $\text{g-C}_3\text{N}_5$, and TiCN composites

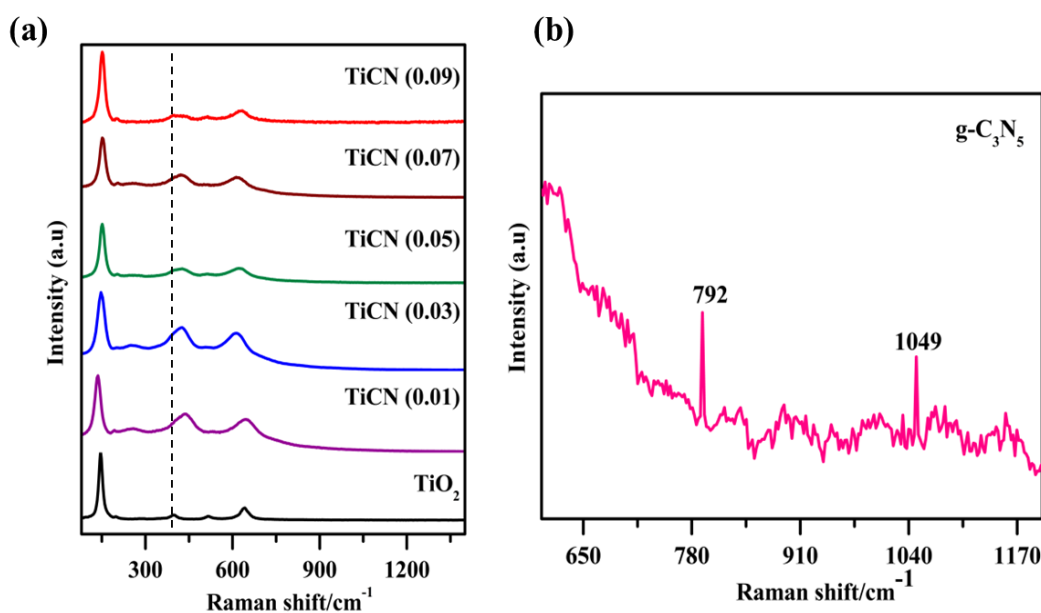


Figure 4. Raman spectra of TiO_2 , $\text{g-C}_3\text{N}_5$, and TiCN composites, highlighting the characteristic vibrational modes and structural features of each sample and their composites

3.3 Raman Analysis

Raman spectroscopy was used to examine the crystalline structure and detect any trace amounts of secondary phases or impurities in the samples, as shown in the Figure 4 [52]. In Figure 4a, the bands observed at 145 cm^{-1} (Eg), 197 cm^{-1} (Eg), 396 cm^{-1} (B1g), 516 cm^{-1} (B1g), and 640 cm^{-1} (Eg) confirm the characteristic properties of TiO_2 anatase [53]. The strong correlation between the Raman and XRD data verifies the high-purity TiO_2 anatase phase prepared through

sol-gel synthesis [54, 55]. Figure. 4b shows bands at 792 cm^{-1} and 1049 cm^{-1} , indicating the presence of the $\text{g-C}_3\text{N}_5$ phase. The Raman spectra for TiCN composites show peaks that are slightly shifted from those of pure TiO_2 . Due to the lower concentration of $\text{g-C}_3\text{N}_5$, the TiCN composite peaks closely resemble the characteristic peaks of pure TiO_2 . These shifts result in peak broadening in the Raman spectra, confirming the presence of both TiO_2 and $\text{g-C}_3\text{N}_5$ in the composite and indicating that the peak shifts arise from changes in

material composition rather than chemical composition. Variations in g-C₃N₅ content influence the structural properties of the composite, as confirmed by Raman spectroscopy, and this data aligns well with XRD. This peak broadening and the observed blue shifts from the positions of TiO₂ suggest increased bonding interactions resulting from structural integration between TiO₂ and g-C₃N₅.

3.4 Scanning Electron Microscopy Analysis

Figure 5 reveals SEM micrographs of uncoated 316LSS and coated with TiO₂, g-C₃N₅ and TiCN coating. In Figure 5A, the SEM image of the uncoated sample reveals prominent grid lines resulting from mechanical polishing of 316LSS [56-58].

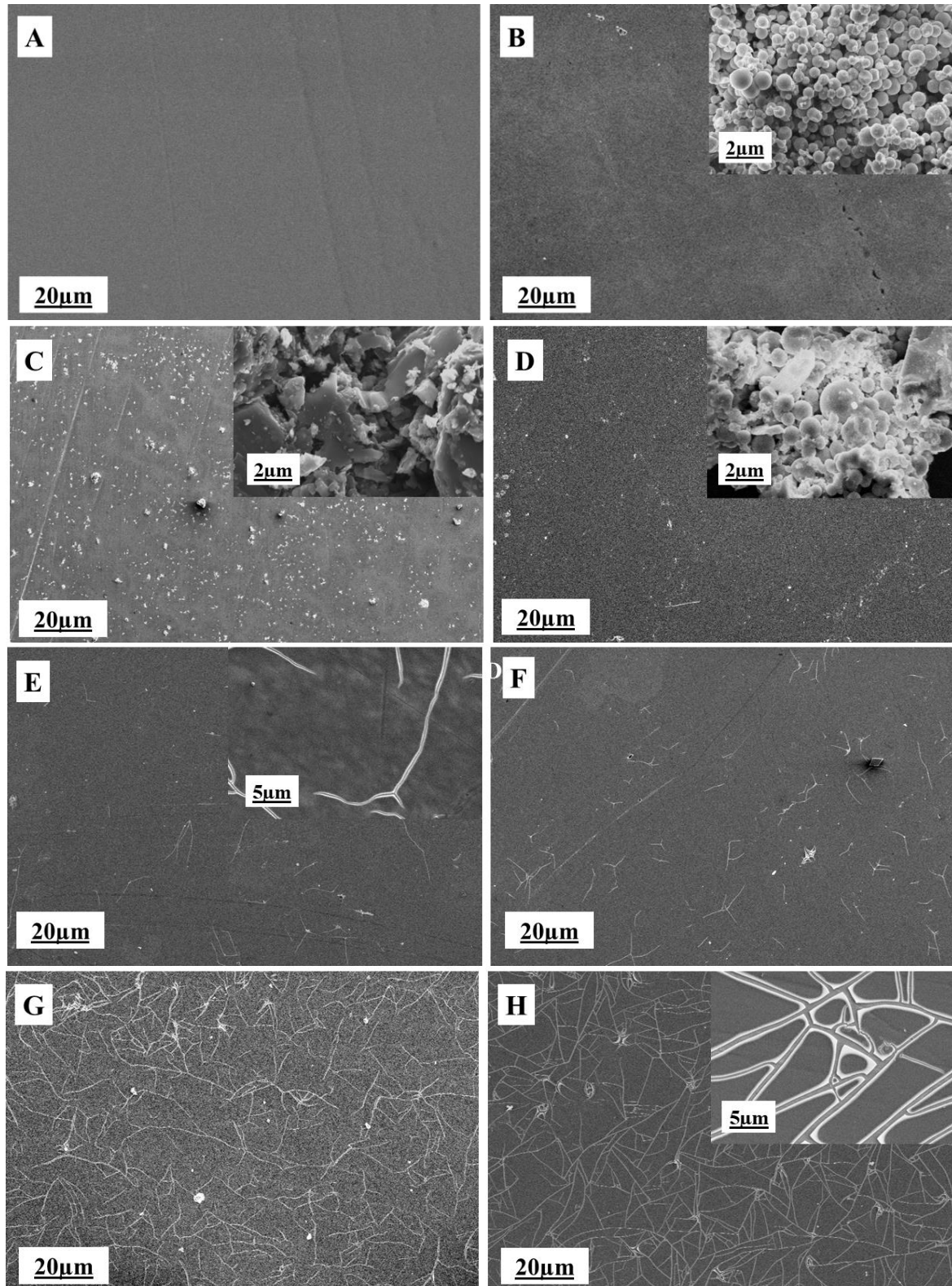


Figure 5. SEM images of A) Bare B) TiO₂ C) g-C₃N₅ D) TiCN (0.01) E) TiCN (0.03) F) TiCN (0.05) G) TiCN (0.07) H) TiCN (0.09), while a magnified view highlights structural detail

Figure 5B shows a TiO₂-coated surface with dense and porous surface morphology, and at magnification, a uniform sphere shape structure was observed [59]. Figure 5C shows the SEM micrograph of the g-C₃N₅-coated sample, showing flat nanosheets with occasional larger agglomerates, and at magnification, stacked sheets-like structures are observed. Figure 5D illustrates the TiCN (0.01) coated sample, which shows a reduced pore size and homogeneous surface and suggests improved coating properties due to the interaction of TiO₂ and g-C₃N₅. These improved surface properties reveal the g-C₃N₅ matrix embedded in the TiO₂. TiCN (0.03) coated sample has enhanced surface properties compared to 0.01 TiCN coating, as shown in Figure. 5E. Beyond the TiCN 0.03 weight percentage, the coating starts to crack due to the thickness of the coating as well as the fast solvent evaporation during the sintering process, which gradually develops the cracks [60-62]. Compared to 0.05, 0.07 and 0.09, weight percentages of TiCN coating have larger cracks, as shown in Figure. 5F, 5G, 5H, which highlights the importance of controlling coating thickness for maintaining mechanical integrity [63]. This uniformity in the coating is crucial to the stability and longevity of orthopedic implants, as it produces a strong mechanical bond between the implant and the surrounding bone tissue [64].

3.5 Atomic force microscopy analysis

Figure 6A shows AFM images of an uncoated 316LSS sample, which has grid lines resulting from mechanical polishing are well corroborated with obtained SEM results. The mean square roughness (Rrms) value of 1.8 nm and an average roughness (Ra) value was 1.3 nm was calculated. Figure 6B shows the TiO₂-coated sample, featuring a uniform spherical

structure that creates a highly porous surface. The TiO₂ coating has a root mean square roughness (Rrms) of 8.8 nm, average roughness (Ra) of 7.1 nm, and average particle size of approximately 50 nm was observed. The g-C₃N₅ coating in Figure. 6C results in flat nanosheets with occasional larger agglomerates adhering on the surface of the sample, which has an average particle size of around 70 – 100 nm. It has a lower surface roughness value Rrms 5.5 nm, Ra 3.2 nm compared to TiO₂ because flat portions are smooth stacked layers, contributing to low roughness. In contrast, agglomerated particles contribute to heightening of the peaks. In addition, Fig. 6D, 6E, 6F, 6G, and 6H display the TiCN composite coating with different weight proportions. It was well-connected sphere shape structures and increased thickness from 0.01 to 0.09 compared to both TiO₂ and g-C₃N₅ coatings, indicating successful incorporation of the g-C₃N₅ into TiO₂ sample and the surface roughness values is TiCN (0.01) Rrms 6.1 nm, Ra 4.8 nm, TiCN(0.03) Rrms 6.7 nm, Ra 5.3 nm, TiCN(0.05) Rrms 7 nm, Ra 5.6 nm, TiCN(0.07) Rrms 7.1 nm, Ra 6.0 nm, TiCN(0.09) Rrms 7.3 nm, Ra 5.6 nm. Higher roughness values suggest a larger pore diameter, which facilitates the incorporation of hydroxyapatite into the pores and improves the bioactivity of the coated surface [65-68]. The surface topography results align well with obtained SEM results.

Figure 7 illustrates the line profile, the significant depth, and the height, indicating that the nanometer to sub-micrometer of the pore's presence was observed. The average particle size of the TiCN composite was around 25-100 nm. This suggests that bioactivity is closely related to the curvature of the particles on the coated film surface [69]. The porous nature of the coating sample promotes bone in-growth, leading to a strong attachment to the bone [70].

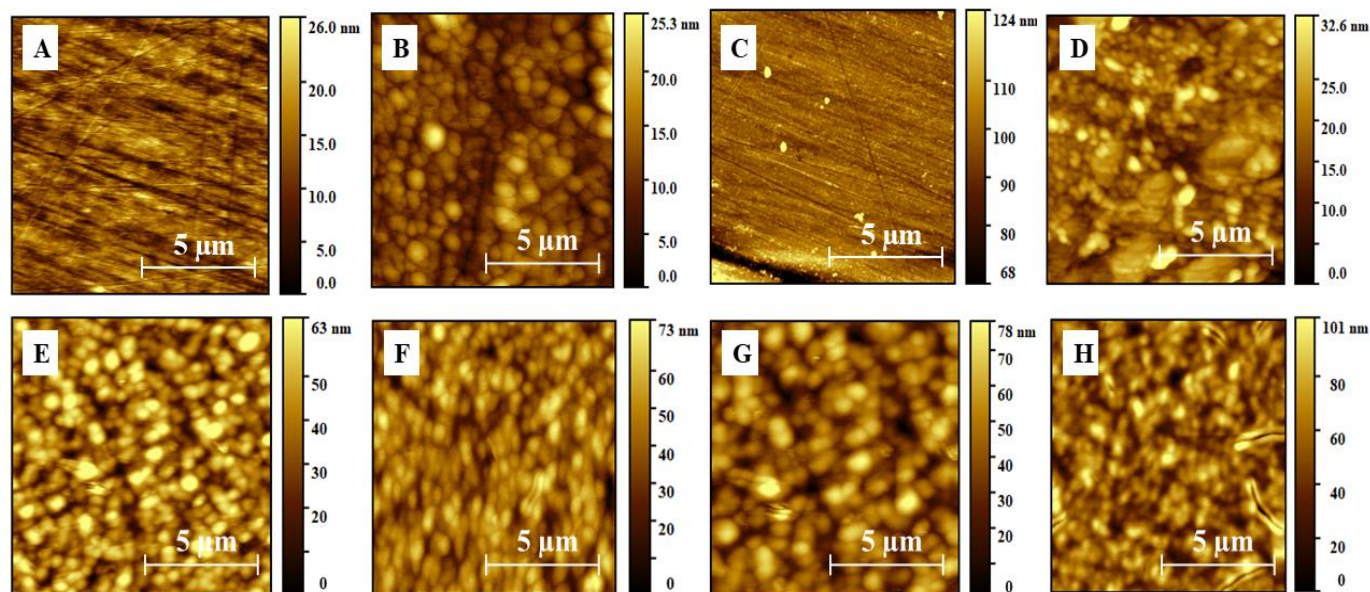


Figure 6. AFM 2D images of A) Bare B) TiO₂ C) g-C₃N₅ D) TiCN (0.01) E) TiCN (0.03) F) TiCN (0.05) G) TiCN (0.07) H) TiCN (0.09) composites

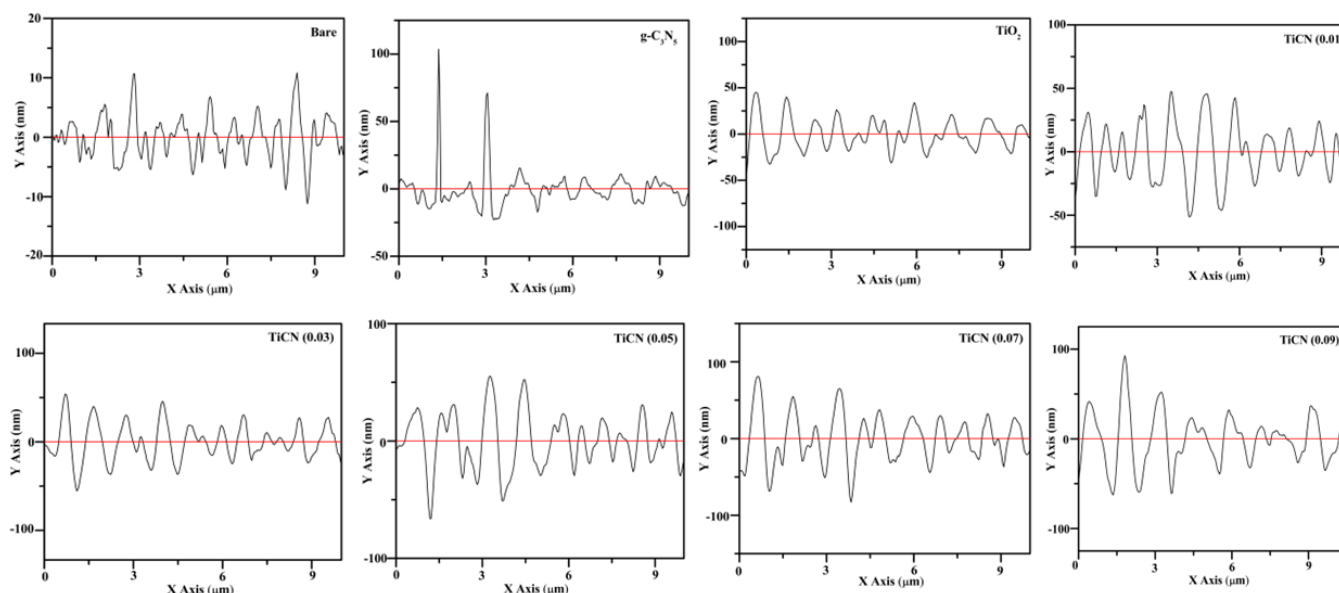


Figure 7. AFM line profile of the uncoated and coated sample, showing the surface roughness and topographical variations across different regions of the sample

These findings suggest that the synergistic interaction between TiO_2 and $\text{g-C}_3\text{N}_5$ in TiCN composite can potentially improve the biocompatibility of coatings.

3.6 Open Circuit Potential

In Figure. 8, OCP measurements showed that the coated samples exhibited higher corrosion resistance than the uncoated 316LSS. The TiO_2 , $\text{g-C}_3\text{N}_5$ and TiCN composite coatings demonstrated a positive shift in OCP values, indicating the formation of protective films. The TiCN (0.03 wt%) composite has the highest OCP value of -0.37 V vs SCE, reflecting superior corrosion resistance. Higher $\text{g-C}_3\text{N}_5$ concentrations (0.05, 0.07, and 0.09 wt%) showed decreased corrosion resistance due to crack formation in the coated film. Therefore, the TiCN (0.03 wt%) composite coating exhibits the enhanced corrosion protection, while the TiO_2 and $\text{g-C}_3\text{N}_5$ coatings showed moderate corrosion protection properties. The positive OCP shift indicates effective corrosion protection [71-73]. The superior corrosion resistance of the TiCN (0.03 wt%) composite coating can be attributed to the optimal concentration of TiCN that promotes the formation of a stable, protective film and avoids structural defects that could compromise the coating's performance. The enhanced OCP shift confirms the positive contribution of TiCN to corrosion protection, while the results from TiO_2 and $\text{g-C}_3\text{N}_5$ coatings demonstrate moderate protection in comparison. These results confirm that adding nitrogen-rich carbon nitride enhances the OCP of the TiCN composite coating.

3.7 Potentiodynamic polarization studies

To investigate the corrosion protection properties of the TiCN sample, a polarization test was conducted, and the corresponding results are presented in Figure 9. The Tafel method was used to extrapolate corrosion potential, corrosion current density and corrosion rate from polarization curves, which are all typically recognized as important parameters for determining corrosion resistance [74, 75]. Generally, the lowest current density and more positive potential indicate better corrosion resistance [76]. For the uncoated 316LSS, the values were determined to be -0.73 V vs SCE -1.9 μA and 0.045 mmpy, respectively. In the case of TiO_2 , the potential -0.57 V vs SCE, current density -2.4 μA and corrosion rate 0.012 mmpy values were obtained, which has enhanced corrosion resistance than bare. For $\text{g-C}_3\text{N}_5$, the absence of a straight-line section confirms the presence of coating. It has a corrosion potential of -0.68 V vs SCE, a current density of -1.7 μA and a corrosion rate of 0.037 mmpy. However, its corrosion rate is almost the same as that of the uncoated 316LSS due to the low adhesive nature of the coating. TiCN (0.03%) composite coated sample exhibited a high positive corrosion potential of -0.35 V vs SCE and low current density of -2.1 μA with a corrosion rate of 0.003 mmpy compared to another TiCN coated sample. The lowest I_{corr} observed for the TiCN coating can be attributed to the strong interfacial interaction between the TiCN coating and the sample, likely due to the high adhesive strength of this coating [77]. The polarization curves bend towards lower current values in the anodic region, indicating that the metal sample has entered a passive state.

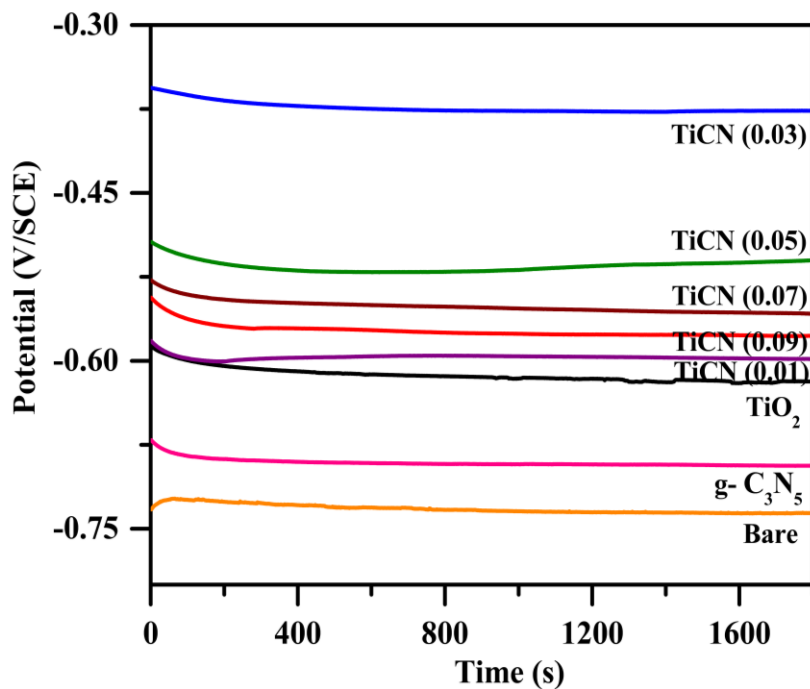


Figure 8. Open circuit potential profiles of uncoated and coated samples

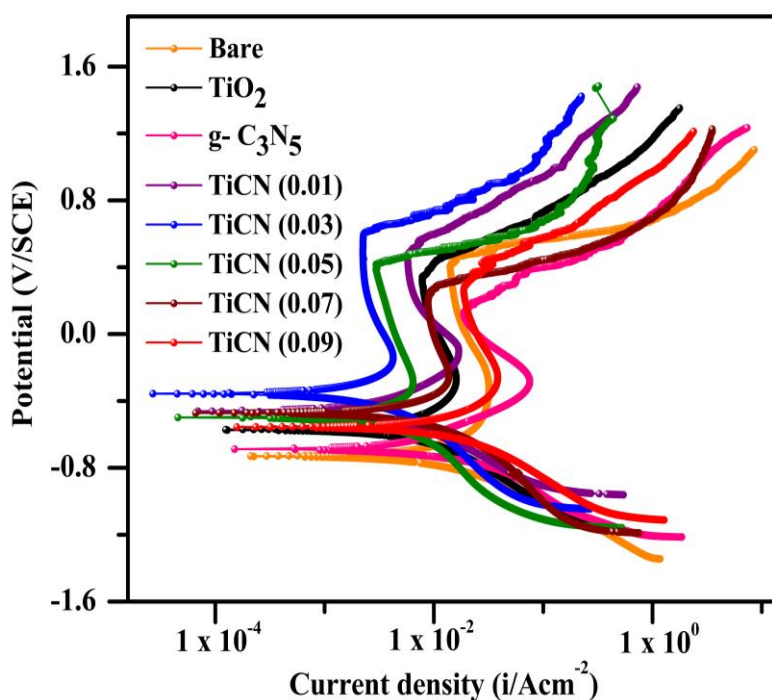


Figure 9. potentiodynamic polarisation graph for uncoated and coated materials

This forms a protective layer like passive film on the metal surface, which significantly slows down the corrosion process [78, 79]. This improvement was due to the TiCN coating on the sample, which hinders the electrolyte solution from reaching the metal sample, leading to a lower current density and increase in corrosion protection efficiency. Potentiodynamic polarization studies confirmed that the TiCN (0.03%) coating has four times higher corrosion potential, significantly improving the corrosion protection performance of the 316LSS sample in the SBF solution.

3.8 Electrochemical impedance spectroscopy

Figure 10A shows the impedance spectra for both coated and uncoated samples, where the uncoated sample exhibits a single semicircle. In contrast, the coated samples showed two semicircles, magnified in Figure. 10B. The initial small semicircle corresponds to the coating layer, with the radius increasing due to the coating. In Figure. 10, the uncoated 316L stainless steel has lower resistance due to the electrolyte penetration occurring through conductive paths in surface pores [80-

82]. However, for the $g\text{-C}_3\text{N}_5$ -incorporated TiO_2 coating, these conductive paths are less accessible due to smaller pore size and higher compactness, which slows electrolyte reach to the sample compared to TiO_2 and $g\text{-C}_3\text{N}_5$. Solution resistance (R_s) depends on the area available for current and the resistivity of the electrolyte, showing minimal variation across samples. An increase in charge transfer resistance (R_{ct}) at the metal/electrolyte interface reduces corrosion rate by slowing electron movement and limiting material deterioration [83]. The TiCN (0.03%) coating shows a high solution resistance ($23 \Omega\text{cm}^2$) and charge transfer resistance ($197 \Omega\text{cm}^2$), suggesting strong corrosion resistance in the electrolyte solution. The equivalent circuits depicted in Figure 11, used for EIS data analysis, incorporate R_s , representing the electrolyte resistance between the working and reference electrodes. R_{ct} and R_1 correspond to the charge transfer resistances for the uncoated and coated samples. The elements Q_{dl} and Q_1 represent the capacitive components associated with the uncoated and coated samples [84, 85]. Surface, which is typically lower for materials that are more prone to corrosion. The increased R_{ct} value in this case is a clear indication that the TiCN (0.03%) coating effectively prevents corrosive ions from interacting with the underlying substrate, making it an excellent barrier. The higher Q_1 values for the TiCN (0.03%) composite suggest that a more stable and compact oxide layer forms compared to the uncoated and other TiCN-coated samples, which contributes to enhanced corrosion protection. Additionally, the TiCN (0.03%) composite coating outperforms other TiCN concentrations in corrosion resistance. This is likely because of the TiCN particles are well-dispersed, facilitating the formation of a dense and continuous protective layer. Therefore, the TiCN (0.03%) composite demonstrates superior corrosion resistance than other TiCN coating.

3.9 In-vitro Characterization

3.9.1 Scanning Electron Microscopy Analysis

Figure 12 shows SEM micrographs of uncoated 316LSS and sample coated with TiO_2 , $g\text{-C}_3\text{N}_5$, and TiCN composites after 14 days of immersion in SBF solution, highlighting differences in hydroxyapatite (HAP) formation across these surfaces. Figure 12A illustrates the SEM micrograph of the uncoated 316LSS sample. Randomly agglomerated spherical particles are observed, signifying the formation of hydroxyapatite (HAP). This growth was attributed to the inherent bioactivity of surgical-grade 316LSS, which promotes HAP deposition due to its favorable interaction with biological fluids [86]. Figure 12B shows the TiO_2 coated sample. TiO_2 encourages HAP nucleation and growth on its surface, contributing to improved bone-cell interaction. Figure 12C displays the $g\text{-C}_3\text{N}_5$ coated sample, where the formation of HAP appears more prominent compared to both the uncoated and the TiO_2 coated sample. This enhanced HAP layer was attributed to the biocompatibility of the $g\text{-C}_3\text{N}_5$ coating, which supports increased HAP nucleation and crystallization [87]. Figure 12D reveals the TiCN composite coating, which exhibits a dense, closely packed spherical shape structure with an interconnected surface, indicating better adhesion with higher bioactivity-induced layers than the other samples. The SBF solution penetrates into the pores of the TiCN coated samples, promoting the formation of a hydroxyapatite-rich layer [88]. It has been reported that bioactive implant materials possessing a bilayer passive film with an inner barrier and outer porous layer encourage better osseointegration and bone tissue integration [89].

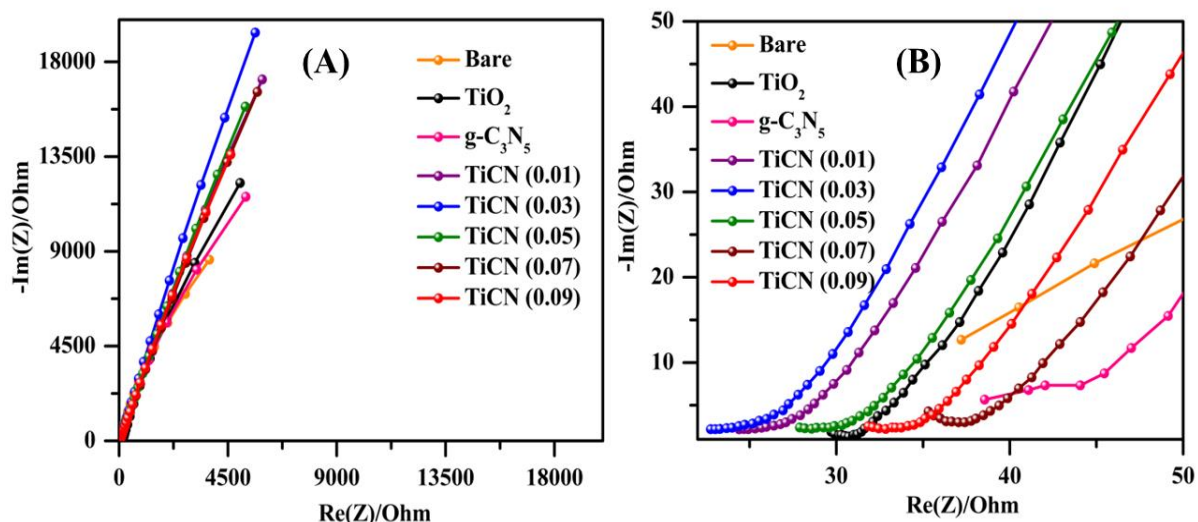


Figure 10. Impedance spectroscopy of A) uncoated and coated sample B) Magnified view of uncoated and coated sample

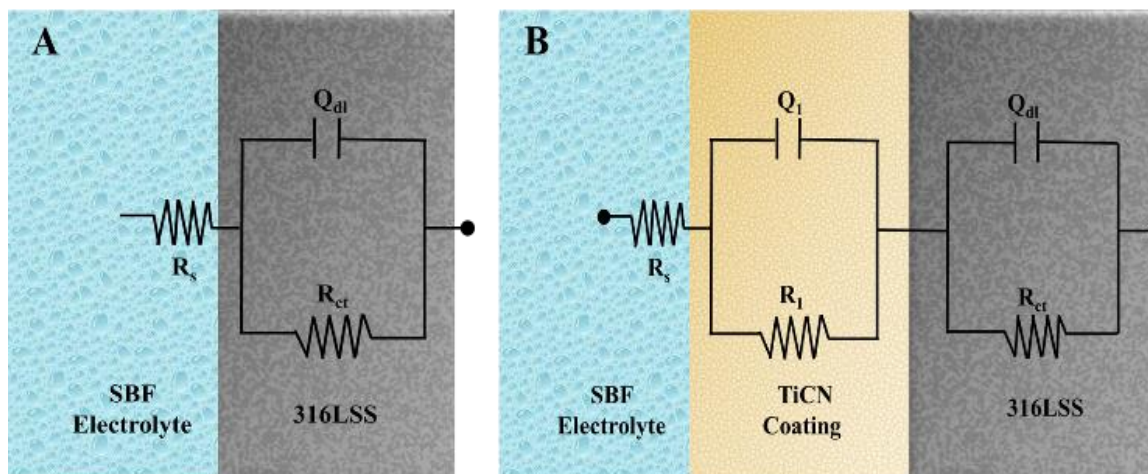


Figure 11. Equivalent circuit diagrams for A) Uncoated 316LSS B) TiCN Coated 316LSS

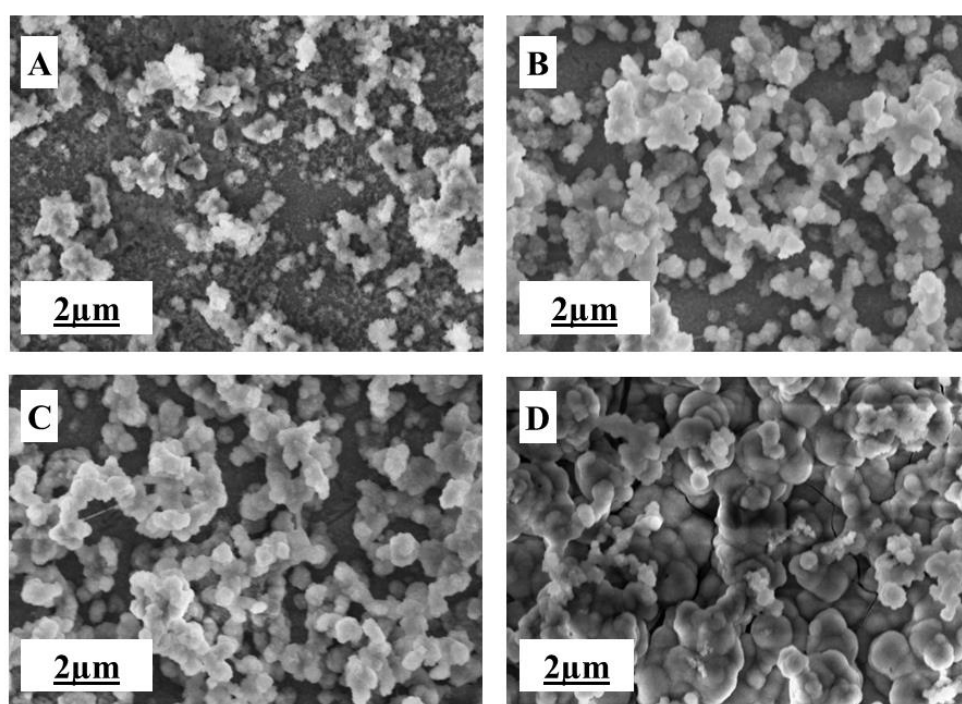


Figure 12. In vitro analysis of the sample: A) Bare B) TiO_2 C) $\text{g-C}_3\text{N}_5$ and D) TiCN (0.03%) composite coating after immersion in simulated body fluid solution

3.10 Raman analysis

Raman spectroscopy is a powerful tool for mineral analysis [90]. The successful deposition of HAP growth was confirmed by the Raman spectrum presented in Figure 13. The characteristic peak of the phosphate ion, associated with the fully symmetric P–O–P stretching mode in HAP, was observed between 954 cm^{-1} and 962 cm^{-1} . The band between 120 cm^{-1} to 300 cm^{-1} corresponds to Ca-PO_4 in a double-bending mode, while the bands from 582 cm^{-1} to 595 cm^{-1} and 611 cm^{-1} to 649 cm^{-1} are attributed to PO_4^{3-} in a triply degenerate bending mode. The peak between 1034 cm^{-1} to 1087 cm^{-1} corresponds to the triply degenerate antisymmetric stretching mode of PO_4^{3-} . The observed Raman bands align well with literature findings [91, 92]. Furthermore, the increased intensity of the TiCN peak

clearly indicates that hydroxyapatite formation significantly enhanced the composites compared to $\text{g-C}_3\text{N}_5$ and TiO_2 coatings. The Raman spectrum clearly shows that $\text{g-C}_3\text{N}_5$ and TiO_2 coatings exhibit fewer peaks related to hydroxyapatite formation than the TiCN composite. The growth of phosphate ions (PO_4^{3-}) on metal surfaces during hydroxyapatite deposition enhances the bioactivity of the implants [93]. The calcium and phosphate ions present in the hydroxyapatite (HAp) improve the biological activity of implants by releasing ions during implantation, which supports bone regeneration [94]. Therefore, the TiCN coating was considered biocompatible due to its tissue bonding characteristics. This indicates that the TiCN coating was more effective for biomedical applications.

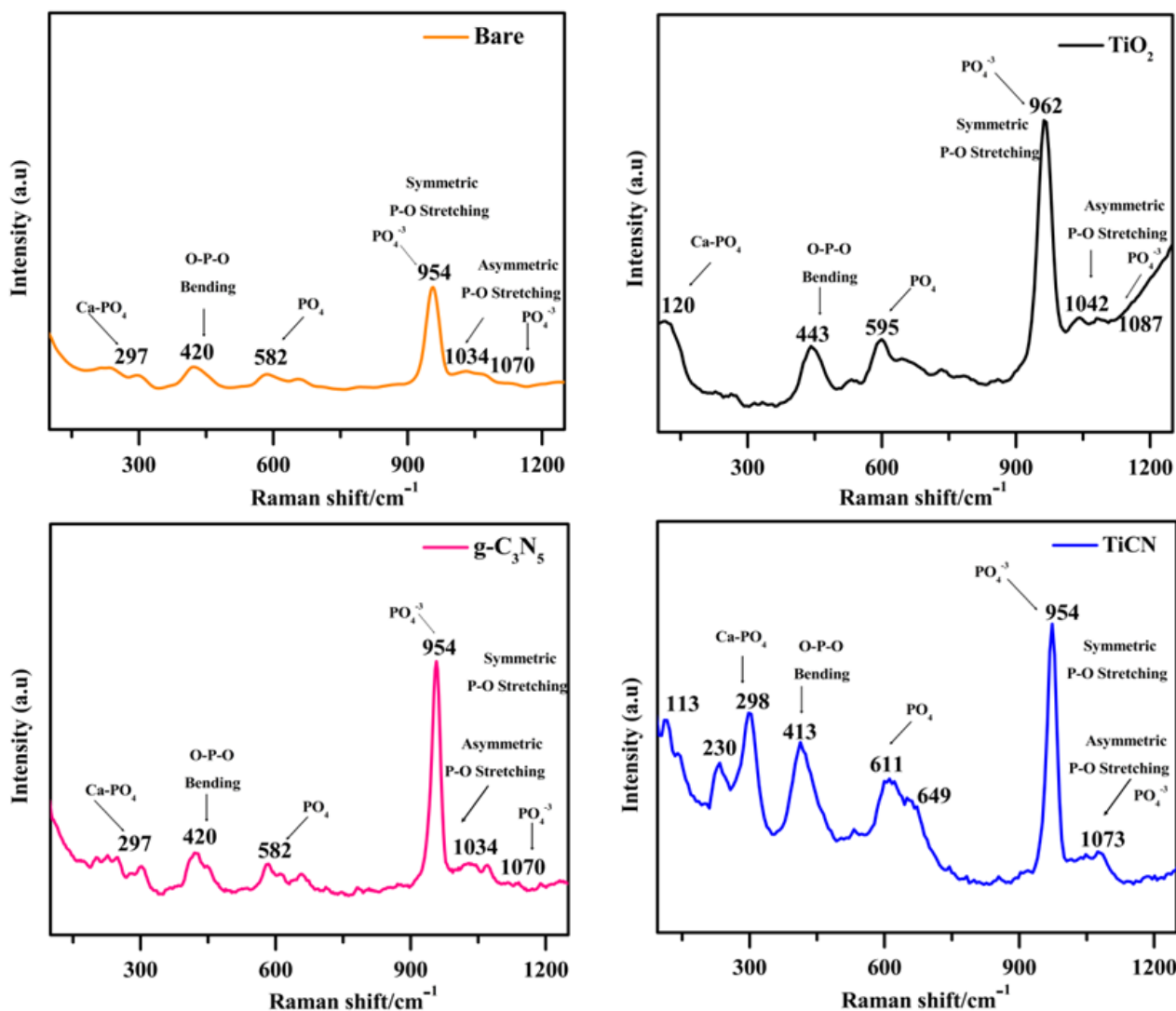


Figure 13. Raman spectrum of HAP deposition on bare, TiO₂, g-C₃N₅, and TiCN (0.03%) composite after the immersion in SBF solution

4. Conclusion

The corrosion performance of the 316LSS sample was assessed after coating with varying weight percentages of TiCN composites. XRD, FT-IR, Raman, AFM, and SEM analyzes confirmed TiO₂ and g-C₃N₅ presence, verifying successful TiCN deposition. Electrochemical analyzes identified the TiCN 0.03% composite as optimal, showing significantly improved corrosion resistance with reduced current density and increased polarisation resistance in SBF solution compared to uncoated and other coated samples. In vitro studies further demonstrated that the TiCN (0.03%) composite supports hydroxyapatite (HAP) growth after 14 days in SBF, with SEM revealing a dense HAP layer for TiCN (0.03%). These findings suggest that the TiCN (0.03%) composite coating was a highly promising protective layer for 316LSS implants, offering superior corrosion resistance and biocompatibility in physiological environments. The TiCN (0.03%) composite coating can be scaled for industrial applications through optimized coating techniques and efficient production processes.

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Supervision. All the authors read and approved the final version of the manuscript.

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Competing Interests

The authors declare that there are no conflicts of interest regarding the publication of this manuscript.

Data Availability

The data supporting the findings of this study can be obtained from the corresponding author upon reasonable request.

Has this article screened for similarity?

Yes

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